	45 min Hybridizatio	on and Bead Preparation	
	Hybridization		
	Prepare 1 - 1,000 ng total RNA in 26 μl.		
	□ Add 4 μl <b>HS •</b> .		
	Add 5 μl <b>PM •</b> , mix until homogeneous.		
	Denature for 5 min at 75 °C / 1,250 rpm.		
	Incubate for 30 min at 60 °C / 1,250 rpm.		
	Bead Washing		
	Resuspend <b>DB</b> •, transfer 75 µl to a fresh tube.		
	Place on magnet for 2 - 5 min, discard supernatant.		
00	Resuspend beads in 75 μl DS •, incubate 2 min on magnet, disconce.	card supernatant. Repeat	
	☐ Resuspend beads in 30 μl <b>DS •</b> .		
	45 min De	epletion and Purification	
		epietion and Porification	
	Depletion		
	Spin down hybridized sample. Add 30 μl of prepared beads. Mix by pipetting 8x, or until homogeneous.		
	Incubate for 15 min at 60 °C / 1,250 rpm. Spin down.		
	Place on magnet for 5 min.		
	Transfer 60 μl supernatant to a fresh tube. <b>ATTENTION:</b> The sup rRNA depleted RNA.	pernatant contains the	
	Purification		
	Add 24 $\mu$ l <b>PB</b> O and 108 $\mu$ l <b>PS</b> O, mix well, incubate for 5 min at	RT.	
	Place on magnet for 5 - 10 min, discard supernatant.		
00	Wash the beads twice with 120 - 150 $\mu$ l 80 % EtOH, 30 sec. <b>ATTENTION:</b> Use 150 $\mu$ l for 1.5 ml tubes.		
	Air dry beads for 5 - 10 min. <b>ATTENTION:</b> do not over dry the beads!		
	Add 12 µl <b>EB</b> O, remove from magnet, mix well, incubate 2 min	Add 12 μl <b>EB</b> O, remove from magnet, mix well, incubate 2 min at RT.	
	Place on magnet for 2 - 5 min, transfer 10 μl of the supernatant stopping point.	to a fresh tube. 🖨 Safe	

**ATTENTION:** Spin down solutions before opening tubes or plates!



	45 min Hybridization and Bead Conditioning		
	Hybridization		
	Prepare 1 - 1,000 ng total RNA in 26 μl.		
	Add 4 µl <b>HS ●</b> .		
	Add 5 μl <b>PM •</b> , mix until homogeneous.		
	Denature for 5 min at 75 °C / 400 rpm.		
	Incubate for 30 min at 60 °C / 400 rpm.		
	Bead Conditioning		
	Resuspend <b>DB</b> ●, transfer 75 µl to a fresh tube.		
	Place on magnet for 2 - 5 min, discard supernatant.		
00	Resuspend beads in 75 $\mu$ l <b>CS •</b> , incubate 2 min on magnet, discard supernatant. Repeat once.		
000	Resuspend beads in 75 µl <b>DS •</b> , place on magnet for 2 - 5 min, discard supernatant. Repeat twice.		
	Resuspend beads in 30 µl <b>DS ●</b> .		
	75 min Depletion and Purification		
	Depletion		
	Spin down hybridized sample. Add 30 µl conditioned beads. Mix by pipetting 8x, or until homogeneous.		
	Incubate for 15 min at 60 °C / 400 rpm. Spin down.		
	Place on magnet for 5 min.		
	Transfer 60 µl supernatant to a fresh tube. <b>ATTENTION:</b> The supernatant contains the rRNA depleted RNA.		
	Purification		
	Add 24 μl <b>PB</b> O and 108 μl <b>PS</b> O, mix well, incubate for 20 min at RT.		
	Place on magnet for 5 - 10 min, discard supernatant.		
	Add 30 µl <b>EB</b> O, remove from magnet, mix well, incubate 2 min at RT.		
	Add 66 μl <b>PS</b> O, mix well, incubate 5 min at RT.		
	Place on magnet for 2 - 5 min, discard supernatant.		
00	Wash the beads twice with 120 - 150 µl 80 % EtOH, 30 sec.		
	Air dry beads for 5 - 10 min. <b>ATTENTION:</b> do not over dry the beads!		
	Add 12 µl <b>EB</b> O, remove from magnet, mix well, incubate 2 min at RT.		
	Place on magnet for 2 - 5 min, transfer 10 ul of the supernatant to a fresh tube.		